Effect of exposure of African catfish (*Clarias batrachus*) to magnetic field on water properties and egg hatching

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Abstract

Magnetic water is produced when water is passed through a magnetic field with the purpose of modifying its structure. The changes in physical and chemical properties of magnetised water affect the biological properties of the organisms. The magnetic field can affect the growth of fish from the embryo to the adult stage. The present study evaluates the effects of magnetic field exposure on water properties and hatchability of the eggs of African catfish (*Clarias gariepinus*). Water was passed through magnetic devices of different intensities; namely: 0.10, 0.15 and 0.20 Tesla. The dissolved oxygen (mgL⁻¹) and pH levels were found to significantly (P ≤ 0.05) increase from 5.92 mgL⁻¹ to 6.33 mgL⁻¹, and from 8.03 to 8.19, respectively. Ammonium (NH₄-N mgL⁻¹) level declined significantly (P ≤ 0.05) (0.20 mgL⁻¹ to 0.16 mgL⁻¹). Salinity (ppt), conductivity (uscm⁻¹) and total dissolved solids (mgL⁻¹) also decreased after magnetization. Significant increase in the rate of hatching was attained in water exposed to a magnetic field of 0.10, 0.15 and 0.20 T. The study demonstrated the benefits of using magnetic devices that are simple, practical and cost-effective.

Keywords: Magnetic field, Magnetized water, African catfish hatchability, Water properties.

Introduction

The African Catfish is a widely consumed fish. Ability of this fish to endure rearing conditions and handling makes it the fish of choice in Malaysian restaurants (Olaleye, 2005). Because the fish can breed easily in captivity, it is a great advantage in farming. Janssen (1985) noted that the artificial reproduction of *C. gariepinus* requires enough flow of suitable quality water. De Graaf et al. (1995) reported that the average hatching percentage of the eggs varied between 28.4 ± 4.5 and 59.1% ± 3.7 for catfish, while Macharia et al. (2005) reported a rate of as low as 4% for the incubated eggs. High rate of fertilization, hatching and survival of larvae at early stage are important factors in the culture of this and other species (Ataguba et al., 2009).

Water quality is determined by variables such as temperature, turbidity, colour, pH, hardness, and the contents of carbon dioxide, unionised ammonia, nitrite and nitrate (Bhatnagar and Devi, 2013). Undesirable water quality can be harmful to aquatic animals. Hence, measures are taken to improve water quality for fish culture.

The magnetic field can affect the growth of fish from the embryo to the adult stage (Chew and Brown, 1989; Tesch et al., 1992; Formicki and Perkowski, 1998; Formicki et al., 2004 a, b; Tanski et al., 2005; Formicki, 2008). It affects the egg’s water uptake after fertilization (Sadowski et al., 2007), motility of developing embryos (Winnicki et al., 2004), embryonic respiration (Formicki et al., 1998), and spatial orientation of embryos and larvae (Formicki et al., 1997). Magnetized (or magnetically treated) water is produced by passing it through a magnetic field of certain strength (Rasoolian et al., 2014). Lund (1947) noted an accelerated growth in plants treated with magnetized water. Since then researchers have used a variety of experimental techniques to study the effects of magnetic field on living organisms. Some studies (Vagria, 1976; Lebkowska, 1991; Baker and Judd, 1996; Gabrielli et al., 2001; Krzemieniewski et al., 2003; Krzemieniewski et al., 2004) have revealed that exposing water to magnetic field influences the water’s surface tension, density, viscosity, hardness, conductivity and solubility of solid matter. These changes in properties affect the biology of the organisms that use the water. According to Sunita and Padmavathi (2013) the high NH₃-N level is detrimental to the fish and thus, a low level of NH₃-N in treatments is desirable for fish growth. The dissolved oxygen is the crucial parameter of water quality because of its effect on metabolism. Likewise, pH affects the metabolism and other physiological processes. The improved hatching of the eggs of Tilapia is due to the effect of magnetic fields on ion fluxes, ligand binding and enhanced distribution and aggregation of intramembranous proteins (Bersani et al., 2004).
The water salinity affects Artemia hatchability (Soundarapandian and Saravanakumar, 2009).

The interaction of magnetic energy with water has stimulated research interest for widening the use of magnetized water in various areas, as well as understanding the fundamental mechanisms of such interactions. Gill et al. (2005) noted significant gaps in knowledge regarding the use of magnetized water in aquaculture. There are still many questions pertaining to this topic in aquaculture, such as the intensity of exposure to magnetic field, effects on water properties, and the effects of these properties on aquaculture organisms and many more. This study aims at studying the effects of exposing water to magnetic field on its properties and hatching of the eggs of African catfish.

Materials and Method

A sequence of procedures was followed to implement the magnetization of water. It involved the usage of a set of magnets with intensities (0.10, 0.15 and 0.20 T). Devices used are shown in Figures 1 and 2.

Figure 1. The magnetization devices of different intensities used in the experiment (a) 0.10 T (b) 0.15 T (c) 0.20 T.

Figure 2. Cross-section of the magnetic devices.

The magnetization of water was carried out at the rate of a single application to the magnetic flux. The professional Plus Multiparameter Water Quality Meter was used to measure water quality parameters. These parameters included water temperature, dissolved oxygen concentration (DO) mg L⁻¹, pH level, specific conductance (SPC) uscm⁻¹, salinity (SAL) ppt, conductivity (CD) uscm⁻¹, total dissolved solids (TDS) mg L⁻¹, ammonia (NH₃-N) mg L⁻¹, and ammonium (NH₄-N) mg L⁻¹. All these parameters were measured in the control (tap water) and magnetized water at intensities of 0.10, 0.15 and 0.20 T. The study was conducted in the hatchery for a week. Magnetizing of water at three different intensities was carried out by allowing it to flow through one of the three devices containing magnets with different field strengths as follows: 0.10, 0.15 and 0.20 T (Figure 3). Water was magnetized by running it through the magnetic devices for two days. Parameters observed were: temperature (ºC), dissolved oxygen concentration, pH level, NH₄-N level, NH₃-N level, SAL, CD, SPC and TDS.

Figure 3. Positions of the magnetic device in the experimental assembly.

Egg incubation

The broodstock was kept separately without feeding after they were injected with 0.35 ml Ovaprim per kg live weight (Oyeleye and Omitogun, 2007). After 12 h, the female brood stock was taken out and held gently in a wet towel while slight pressure was applied on the abdomen to release the eggs into a dry bowl as suggested by Hogendoorn (1979). The male was sacrificed to obtain the gonads which contained the milt. The mixture of eggs and milt was stirred gently for about 1.0 - 2.0 minutes to allow fertilization to take place. Within a few minutes, the eggs absorbed water and became sticky. Sixteen incubation/hatching tanks were used in this study, each containing 18 L of freshwater per tank. The experiment was conducted in four replications. The eggs were dispersed in the hatching tanks by a pipette. About 5 ml of egg-containing sample was incubated in each tank. For reference, 1ml of egg is equivalent to about 644 eggs (each tank has 3220 egg). The water was aerated and the experiment was conducted at the temperature 26.2 - 26.38°C.

Hatching percentage was calculated for each treatment system. The fertilized egg masses were incubated in the spawning tanks for two days. The percentage of hatchlings that emerged following the incubation was estimated over days after fertilization. The density of larvae (number of larvae per tank) was determined in 5 samples (100 ml per sample). The number of larvae was counted in a sample of 100 m drawn from each tank.

\[
\text{Hatching rate} = \frac{\text{No. of eggs hatched}}{\text{Total no. of eggs in a batch}} \times 100 \ldots \text{Eq. (1)}
\]
The survival rate was calculated over seven days by using the equation (2):

\[
\text{Survival rate} = \frac{\text{No. of live hatchling of eggs to larval stage}}{\text{Total no. of hatching eggs}} \times 100 \ldots \text{Eq. (2)}
\]

Statistical software SAS 9.0 was used for the one-way analysis of variance (ANOVA) and Duncan’s multiple test was applied to determine the significance of differences among the water properties, hatchability rate and survival rate of African catfish at magnetic intensities of 0.10, 0.15 and 0.20 T. The differences are regarded as significant if \( P \leq 0.05 \).

Results and Discussion

Effect of exposure to magnetic field on water properties

Generally, the dissolved oxygen increases with increasing water magnetization. The best result shows an increase of 0.41 mg L\(^{-1}\) after the water was magnetized with field strength of 0.20 T. Constant aeration was necessary to keep the eggs in suspension and to provide sufficient dissolved oxygen to the eggs to aid in their hatching. A minimum of three parts per million dissolved oxygen during the incubation is crucial for the metabolism of the catfish. This result is in agreement with Al-Ibady (2015) who observed that the increase in magnetic intensity led to significant increase in dissolved oxygen concentration compared to normal water (control).

It can be seen from the data that the pH increased slightly with magnetization intensity. The results show a maximum increase of 0.16 pH after the water was magnetized with field strength of 0.20 T. The exposure of water to magnetic field softens the water and increases the pH (Lowe, 1996). This is in agreement with Hasaani et al. (2015) although the authors reported a 12% increase in water pH after magnetization. The effect depends on the intensity of the magnetic field exposure (AbdelTawab et al., 2011).

Results of these treatments show a significant decrease in NH\(_4\)-N concentration compared to normal water. The lowest decrease of NH\(_4\)-N was recorded from 0.20 to 0.16 mg L\(^{-1}\) after the water was magnetized with field strength of 0.20 T. The magnetic field increased the free radical formation, while the reactivity and oxidation potential of those chemical compounds may have reduced the concentration of organic matter contained in the analyzed liquids (Krzemieniowski et al., 2002).

The water salinity showed no significant effect on the trend with magnetization intensity. Brownie and Wanigasekera (2000) and Vanhaecke and Sorgeloos (1989) reported that using high intensity of magnetic field lead to decrease in the water salinity.

The total dissolved solids showed no decreasing trend with magnetization intensity. These results are in agreement with those reported by Gilani et al. (2014), Vanhaecke et al. (1984) and Hasaani et al. (2015). Researchers have demonstrated that exposure to magnetic field affects the molecular and physico-chemical properties of water by altering the water nucleus (Hasson and Bramson, 1985; Gehrl et al., 1995; Coey and Cass, 2000; Cai, et al., 2009). The water molecules align in one direction due to the relaxation of bonds, and decrease in their angle to below 105º (Lowe, 1996). The consolidation degree between water molecules decreases which could cause an increase in the size of molecules. This can change the pH and TDS properties in the water. The results of the current study are in disagreement with Al-Ibady (2015), who observed an increase in the total dissolved solids with increase in exposure to magnetic field intensity. The effect of magnetic water treatment depends on the composition of the treated water, magnetic field strength, rate of water movement, duration of its stay in the magnetic field and other factors. The reason for the increased solubility of the salts when exposed to the magnetic fields is due to the magnetically modified chemical and physical properties of water. Results of water properties in the experimental and control sets are shown in Table 1.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Normal water (control)</th>
<th>0.10 T</th>
<th>0.15 T</th>
<th>0.20 T</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH</td>
<td>8.03 ± 0.03 c</td>
<td>8.13 ± 0.02 b</td>
<td>8.17 ± 0.01 a</td>
<td>8.19 ± 0.01 a</td>
</tr>
<tr>
<td>SPC uscm(^{-1})</td>
<td>242.3 ± 4.60 b</td>
<td>240.5 ± 0.21 a</td>
<td>240.3 ± 0.17 a</td>
<td>240.2 ± 0.26 a</td>
</tr>
<tr>
<td>CD uscm(^{-1})</td>
<td>247.55 ± 3.71 a</td>
<td>246.75 ± 0.52 a</td>
<td>246.05 ± 0.62 a</td>
<td>246.30 ± 0.40 a</td>
</tr>
<tr>
<td>TDS mg.l(^{-1})</td>
<td>157.3 ± 1.71 b</td>
<td>156.3 ± 0.38 a</td>
<td>156 ± 0.00 a</td>
<td>156 ± 0.00 a</td>
</tr>
<tr>
<td>SAL/ ppt</td>
<td>0.113 ± 0.01 a</td>
<td>0.11 ± 0.00 b</td>
<td>0.11 ± 0.00 b</td>
<td>0.11 ± 0.00 a</td>
</tr>
<tr>
<td>NH(_4)-N mgL(^{-1})</td>
<td>0.20 ± 0.03 a</td>
<td>0.17 ± 0.01 b</td>
<td>0.17 ± 0.01 b</td>
<td>0.16 ± 0.01 b</td>
</tr>
<tr>
<td>NH(_3)-NmgL(^{-1})</td>
<td>0.02 ± 0.02 ±</td>
<td>0.02 ± 0.02 ±</td>
<td>0.02 ± 0.02 ±</td>
<td>0.02 ± 0.02 ±</td>
</tr>
<tr>
<td>DO mgL(^{-1})</td>
<td>5.92 ± 0.07 c</td>
<td>5.92 ± 0.07 c</td>
<td>6.11 ± 0.08 b</td>
<td>6.33 ± 0.04 a</td>
</tr>
<tr>
<td>C(^\circ)</td>
<td>26.20 ± 0.06 a</td>
<td>26.35 ± 0.00 ±</td>
<td>26.40 ± 0.00 ±</td>
<td>26.38 ± 0.00 ±</td>
</tr>
</tbody>
</table>

*Same superscripts in rows are not significantly different according to Duncan’s test if \( P \leq 0.05 \).*

Effect of exposure to magnetic field on hatching percentage

The exposure of African catfish to magnetic field has a significant effect (\( P \leq 0.05 \)) on egg hatchability. Exposure to 0.10, 0.15 and 0.20 T achieved the hatchability rate of 55.03, 59.22 and 66.76%, respectively, compared with the hatching rate 51.68% in the normal water. Environmental factors that may affect egg and sperm quality in fish include the diet of the brood fish and the physiochemical conditions of the water in which the eggs are incubated (temperature, salinity, etc.)
and pH of the water). The hatching process is dependent upon water temperature while the hatching rate is depends on the water quality, oxygen level, temperature, and water hardness (Megbowon et al., 2013). The hatching rate is higher for magnetized water treatments of 0.10, 0.15 and 0.20 T, than the control group (Figure 4). According Formicki et al. (2015), exposure to magnetic field positively affected the fish sperm motility and increased percentage of fertilized eggs by 12.7% for 1 mT, 15% for 10 mT, 11.4 for 3 mT, and 22.1 for 1 mT compared to the control. Egg fertilization rates of spermatozoa held for 24 hours under effect magnetic field intensity of 1 mT was: 71.32% at 5 mT and 59.99% under the intensity of 10 mT. In the control sample, egg fertilization rate was only 32.60% (Formicki et al., 2013).

The magnetic field stimulates the breathing process of embryonic tilapia, thereby enhancing their morphogenesis and thus gastrulation and closing of the blastopore, during organogenesis and development of the blood circulation system (Formicki and Winnicki, 1998).

Table 2 summarizes the effect of magnetized water on the hatching percentage of the catfish. Increases of 6.09%, 12.74%, and 22.59% in the hatching rate were attained in water exposed to magnetic field of 0.10, 0.15 and 0.20 T, respectively. The best average hatching result was attained when the water was magnetized with field strength of 0.20 T.

<table>
<thead>
<tr>
<th>Magnetic field strength</th>
<th>Increase of hatching percentage</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.10 T (%)</td>
<td>6.09†</td>
</tr>
<tr>
<td>0.15 T (%)</td>
<td>12.74†</td>
</tr>
<tr>
<td>0.20 T (%)</td>
<td>22.59†</td>
</tr>
</tbody>
</table>

The statistical analysis refers to the effect of magnetized water on the hatching and surviving larvae of the catfish. It can be seen that increase in hatchability is significant (P ≤ 0.05) when the water has been magnetized at 0.20 T, as shown in Table 3.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Treatment</th>
<th>Number of hatchlings</th>
<th>Number of surviving larvae</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
<td>6662.00 ± 616.89</td>
<td>3327.80 ± 289.10</td>
</tr>
<tr>
<td></td>
<td>0.10 T</td>
<td>7092.00 ± 296.86</td>
<td>3510.00 ± 180.00</td>
</tr>
<tr>
<td></td>
<td>0.15 T</td>
<td>7632.00 ± 1004.56</td>
<td>3401.80 ± 632.72</td>
</tr>
<tr>
<td></td>
<td>0.20 T</td>
<td>8604.00 ± 756.28</td>
<td>4193.80 ± 564.45</td>
</tr>
</tbody>
</table>

*Same letters in column are not significantly different according to Duncan’s test (P ≤ 0.05).

According to Tyari et al. (2014), magnetic water increases the solubility of minerals which eventually improves the transfer of nutrients to the fish. It can be assumed that improved hatchability of the eggs of C. gariepinus is due to the magnetic field influencing the membrane functions through the local effect on ion fluxes, ligand binding and alteration in the distribution and aggregation of the intramembranous proteins (Bersani, 1997).

**Conclusion**

Treatments involving the effect of magnetic field of the strengths 0.10, 0.15 and 0.20 T have demonstrated certain variations in some of the physical and chemical properties of water such as DO (mg L⁻¹), pH, NH₄-N, SPC (uscm⁻¹), CD (uscm⁻¹), and TDS (mg L⁻¹). Significant increase from 51.68 to 55.03, 51.68 to 59.22 and 51.68 to 66.67 in African catfish.
hatchability rate was attained in water exposed to a magnetic field of 0.10, 0.15 and 0.20 T, respectively. There was only minor effect of exposure to magnetic field on the survival rate of the fish. The most significant hatching rate of 66.76% was attained when the water was exposed to field strength of 0.20 T compared to the control where it was 51.68%. Results of this study have positive implications for aquaculture because the higher rate of hatching will increase and economize the fish seed supply for aquaculture.

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