

REVIEW ARTICLE

Apoptosis: Dual Role in Aetiology and Cure

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ABSTRACT

Apoptosis is a programmed cell death which occurs following a variety of stimuli. Physiologically the process is important for morphogenesis of organs and homeostasis of different types of cells. Apoptotic cell death is responsible for a variety of pathologic states such as elimination of cell death in mutated cells, infected cells, tumour cells and transplant rejection well as the pathological atrophy. In this review, there is discussion about the control of apoptosis, detection methods of apoptosis, its association with infectious and non-communicable diseases. Intracellular microorganisms survive through inhibition of host cell apoptosis as well as they destroy the parenchymal cells causing impaired functions. It plays important role in tumourigenesis. There are possible therapeutic roles of drugs that modify apoptosis in human diseases.

INTRODUCTION

Apoptosis is a process of programmed cell death and the word was derived from Greek word meaning "falling off" which is an analogy to leaves falling off trees¹. Apoptosis follows a specific stimulus and stimulated by rapid activation of endonucleases²⁻⁴. It is a physiological process occurring during organogenesis in embryonic life, tissue homeostasis such as atrophy of cells in senility, shrinkage of breast and reproductive organs after reproductive period. It has a role in some pathological conditions such as removal of DNA damaged cells and immune cells on removal of cytokines and growth factors^{5,6}.

Recent studies found that some DNA lesions such as base-alkylations, DNA cross-links and DNA-double-strand breaks (DSBS) can trigger apoptosis⁷. Programmed cell death is initiated by activations of endonucleases which causes proteolysis of DNA repair proteins, cytoskeletal proteins, and the inhibitor of caspase-activated deoxyribonuclease^{8,9}.

Detection of Apoptosis

Histologically apoptotic cells are characterized under light microscope by cell shrinkage, blebbing of plasma membrane, small fragmentation of cell cytoplasm containing pyknotic remnants of nuclei. It can also be detected by terminal deoxynucleotidyltransferase (TdT) mediated dUTP-biotin nick end labelling (TUNEL) method to see the nuclear fragmentation¹⁰ which is based on specific binding of TdT to 3'-OH ends of DNA of nuclei undergoing apoptosis on the histological sections or cell preparation in flow or laser scanning cytometry^{10,11}. Degradation of genomic DNA results in formation of DNA fragments which on agarose electrophoresis show ladder DNA¹². Ligation-mediated polymerase chain reaction (LMPCR) can amplify DNA fragmentation specific to apoptotic cell death^{13,14}. Ultrastructurally, nuclear chromatin condensation, cells break up into membrane-bound bodies, apoptotic bodies, which are phagocytosed and degraded by cells such as histiocytes, endothelial cells, epithelial cells or neoplastic epithelial cells^{1,13}. Oligonucleotide and protein microarray methods detect the genes and proteins that regulate apoptosis through the Fas system^{15,16}.

Control of Apoptosis

Apoptosis is initiated by the extrinsic pathway, which can be triggered by ligation of death receptors which activates caspase-8 and the intrinsic pathway initiated by cellular stress by activation of caspase-9 following the cellular stress or the granzyme B pathway, which uses granzyme B to kill the target cells¹⁷. Double-

strand specific caspase is activated and cleaves the chromosomal DNA into large fragments of 50 – 300 kb¹⁸. Some studies describe about the caspase independent processes which requires apoptosis inducing factor (AIF)¹⁷. There are many genes identified in recent years which are responsible for apoptosis²⁰. *Bcl2* oncogene promotes cell survival by blocking programmed cell death that prevents the permeability of mitochondrial outer membrane²¹. Mutation of transmembrane receptor protein, CD95/Fas, that interact with Fas associated death domain (FADD) to recruit pro-caspase 8²². Bax which is a bcl2 homologous protein when stimulated inserts into the mitochondrial outer membrane to release cytochrome C in apoptosis. P53 has a major role in inducing apoptosis: it regulates cell cycles by allowing damaged DNA to repair, inducing Bax expression²³, through activation of Noxa²⁴, increasing the level of reactive oxygen radicals (ROS), all of which can trigger release of cytochrome C. Inhibitors of apoptosis proteins (IAPs) are composed of 70 amino acids termed BIR (baculoviral IAP repeat) discovered in genome of baculoviruses²⁵. Cyclin D1 promotes cell proliferation and inhibits drug-induced apoptosis²⁶. Apoptotic peptidase activating factor-1 (Apaf-1) is responsible for initiating apoptosis downstream mitochondrial damages. Loss of expression of Apaf-1 is associated with recurrences of cancer²⁷.

Apoptosis and Non-communicable Diseases

Apoptosis is a very important process in development of mammals. It deletes some organs during organogenesis, controls the number of cells, elimination of potentially dangerous cells. Mutation of FANCC, which has effects on the failure of apoptosis of haemopoietic stem cells (HSC) on exposure to growth factor deprivation causing DNA damage, is described in Fanconi anaemia (FA)²⁸. Defective Fas-induced apoptosis can lead to autoimmune lymphoproliferative syndrome in human²⁹ which present with

reactive lymphadenopathies and autoimmune manifestations. Apoptosis plays a role in pathogenesis of gout. Delayed spontaneous and TNF- α induced apoptosis is observed in acute gouty arthritis³⁰ and renal damage is caused by apoptosis of renal tubules induced by uric acid³¹. Mutation of p53, a gene that blocks the damaged cell to enter cell cycle and directs towards apoptosis, leads to genetic instability characteristic of many cancer cells². Apoptosis and the genes that control it have a significant effect on the carcinogenesis in the steps of initiation and progression, tumour biological behaviour such as metastasis and the morphologic types of cancer²². Mutation of caspase-8 gene is associated with advanced gastric cancer, hepatocellular carcinoma³² leukaemia³³ and medulloblastoma³⁴. Bcl-2, a protein that blocks apoptosis, is associated with B cell lymphoma and inhibitors of apoptosis proteins (IAPs) is found in MALT lymphoma³⁵. Massive and selective apoptosis lead to neurodegenerative diseases and neural tube defects³⁶. Mutation of neuronal apoptosis inhibitory protein (NAIP) was shown in spinal muscular atrophy³⁷. Apoptosis of cardiomyocytes induced by non-encoding RNAMeg3 is associated with myocardial infarction and biomarkers of cell death such as MAC were studied in post-mortem cases in an attempt to identify the new diagnostic markers^{38, 39}.

Apoptosis and Infections

Bacteria-induced host cell death by apoptosis to enhance their replication and survival⁴⁰. Intracellular organisms such as *L. pneumophila*, and *Chlamydia* spp. survive in their host cells by inhibiting the macrophage apoptosis⁴¹. A mutant gene *nuoG* in *M. tuberculosis* decreased the ability to inhibit macrophage apoptosis and subsequently reduced the virulence of the organism⁴². Bacterial infection of central nervous system with group B Streptococcus lead to apoptosis of neurons in the presence of microgli⁴³. Regeneration of neural tissue occurs after injury, but it is not sufficient to

replenish the cells required for normal function. Apoptosis and necroptosis of neurons and glial cells occur in meningitis and it is responsible for neurological complications⁴³. HIV 1 virus evades macrophage apoptosis by transcription of prosurvival genes via MAP2K1/ERK2 pathway⁴⁴. Placental malaria usually complicated by poor neonatal outcome due to growth retardation is related to apoptosis-related mechanism⁴⁵. Hepatitis B and C viruses cause chronic liver disease and parenchymal damage by escaping viral clearance by apoptosis⁴⁶. Levels of Caspase-3, which triggers apoptosis, are low in *Mycobacterium tuberculosis* infected pulmonary alveolar macrophages rendering them to escape from apoptosis and create a more favourable environment for intracellular growth of bacteria⁴⁷. Apoptosis is both qualitatively and quantitatively increased in lepra type 1 reaction and proposed to be one of the variables for high detection of the reaction⁵⁹ and lepromatous leprosy⁴⁸. High level of nerve growth factor (NGF), a neurotrophin that causes apoptosis of Schwann cells and nerve damage, is correlated with nerve damage⁴⁹. Pathogens like *Leptospira interrogans* escape phagocytosis by rendering apoptosis of macrophages through Fas-FasL/Caspase-8/-3 pathway⁴⁹.

Application in Therapeutics

Apoptosis is involved in tumourigenesis and other human diseases, it is also a centre of target for treatment. Inhibitors of apoptosis such as IAP may serve as a molecular target for specific anticancer therapy. IAPs can also be used as an indicator of prognosis after therapy as highly expressed in tumour cells⁵⁰. IAP counter-reacts the high basal caspase activity of tumour cells selectively^{51, 52}. Survivin, a member of IAP, is implicated management of cancer in different aspects such as targeting the tumorigenesis, neovascular angiogenesis, and immunotherapy target. Recombinant survivin protein used as a immunotherapy was effective as an adjuvant therapy in human melanoma cells^{53, 54}. Some advances have been made to focus on TNF-related

apoptosis-inducing ligand (TRAIL) for cancer treatment⁵⁵. Agonistic antibodies to TRAIL receptors can induce apoptosis of the tumour cells. Antiapoptotic- Bcl-2 family proteins are implicated in indicators of chemotherapy resistance in prostate cancer and other solid cancers and haematological malignancies⁵⁶. Parenchymal cell death in diseases is a major concern in treating many diseases. Caspase-8, after activation by caspase cascade, is responsible for tumour cell motility by acting on focal adhesion complexes⁵⁷. Evidence showed that pan-caspase inhibitors have profound effect on acute TNF- and Fas/FasL mediated apoptosis in hepatocytes and caspase mediated metastasis⁵⁷. Short term uses of inhibitors of apoptosis are implicated in conditions such as ischaemic stroke, spinal cord injury, reperfusion injury and organ transplant. Arthritis, Alzheimer's disease, and Huntington's disease may benefit from their long-term usage⁵⁸. Anti-angiogenic cancer therapy targets the vascular endothelial cells by inducing the Fas-ligand and initiating the apoptosis cascade⁵⁹. Understanding the underlying mechanisms such as MAP2K1/ERK2 pathway and host cell K⁺ channels inhibition helps to design effective antiviral drugs for HIV and HCV^{44,46}.

CONCLUSION

Apoptosis, programmed cell death, is an essential process in development, control of immune system and tissue homeostasis in living. In addition to its physiological functions, it also plays a role in aetiology of tumour formation and in the mechanism to reduce tumour cell burden in cancer caused by apoptosis of tumour cells. However, there are still some controversial theories on the use of apoptosis modifying therapeutics. Attempts should be made to consider several therapeutic aspects of apoptosis to improve the patient's outcome.

CONFLICT OF INTEREST

The authors declare that they have no competing interests in publishing this article.

REFERENCES

1. Kerr JF, Wyllie AH, Currie AR. (1972). Apoptosis: A basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br J Cancer* 26 (4): 239 – 257.
2. Albert B. (2002). The molecular basis of cancer cell behavior, in *Molecular Biology of the cell* (New York - Garland Science). Retrieved from [https:// www.ncbi.nlm.nih.gov/ books/NBK26902/](https://www.ncbi.nlm.nih.gov/books/NBK26902/)
3. Vaux DL. (2002). Cell death and differentiation. *Apoptosis Timeline* 9: 349 – 354.
4. Arends MJ, Morris RG, Wyllie AH. (1990). Apoptosis. The role of the endonuclease. *Am J Pathol* 136 (3): 593 – 608.
5. Alaa El-Din HS, Afaf IE, Ekbal TW. (2015). Histological and histochemical studies on the early developmental stages of the Egyptian Toad *Bufo regularis* Reuss. *Open Journal of Animal Sciences* 5: 142 – 156. doi: 10.4236/ojas.2015.5
6. Wynn AA, Kazuhisa M, Emi M et al. (2001). Role of granulocyte/macrophage colony-stimulating factor in zymocel-induced hepatic granuloma formation. *Am J Pathol* 158 (1): 131 – 145. doi: 10.1016/S0002-9440(10)63951-X.
7. Roos WP, Kaina B. (2006). DNA damage-induced cell death by apoptosis. *Trends in Molecular Medicine* 12 (9): 440 – 450.
8. Zhang X, Yaming C, Larry WJ et al. (2005). Bench-to-bedside review: Apoptosis/programmed cell death triggered by traumatic brain injury. *Crit Care* 9 (1), 66 – 75. doi: 10.1186/cc2950
9. Clark RS, Kochanek P, Watkins SC et al. (2000). Caspase 3 mediated neuronal death after traumatic brain injury in rats. *Journal of Neurochemistry* 74 (2): 740 – 753.
10. Fayzullina S, Martin LJ. (2014). Detection and analysis of DNA damage in mouse skeletal muscle in situ using the TUNEL Method. *J Vis Exp* (94): 1 – 9. e52211, doi: 10.3791/52211.
11. Darzynkiewicz Z, Galkowski D, Zhao H. (2008). Analysis of apoptosis by cytometry using TUNEL assay. *Methods* 44 (3): 250 – 254.
12. Ueda N, Shah SV. (1992). Endonuclease-induced DNA damage and cell death in oxidant injury to renal tubular epithelial cells. *J Clin Invest* 90: 2593 – 2597.

13. Wyllie AH. (1972). Death in normal and neoplastic cells. *J Clin Path* 27 (7): 35 – 44.
14. Stanley K, Blaschke A, Chun J. (1997). Apoptotic DNA fragmentation is detected by a semi-quantitative ligation-mediated PCR of blunt DNA ends. *Cell Death Differ* 4: 66 – 75.
15. Hofmann WK, Vos S, Tsukasaki K et al. (2001). Altered apoptotic pathways in mantle cell lymphoma detected by oligonucleotide microarray. *Blood* 98: 787 – 794. doi: 10.1182/blood.V98.3.78.
16. Matei C, Tampa M, Caruntu C et al. (2014). Protein microarray for complex apoptosis monitoring of dysplastic oral keratinocytes in experimental photodynamic therapy. *Biol Res* 47 (1): 33; 1 – 19. doi: 10.1186/0717-6287-47-33
17. Ghavami S, Hashemi M, Ande SR et al. (2009). Apoptosis and cancer: Mutations within caspase genes. *J Med Genet* 46: 497 – 510. doi: 10.1136/jmg.2009.066944.
18. Oberhammer F, Wilson JW, Dive C et al. (1993). Apoptotic death in epithelial cells: Cleavage of DNA to 300 and/or 50 kb fragments prior to or in the absence of internucleosomal fragmentation. *EMBO J* 12 (3): 3679 – 3684.
19. Susin SA, Lorenzo HK, Zamzami N et al. (1999). Molecular characterization of mitochondrial apoptosis-inducing factor. *Nature* 397: 441 – 446.
20. Müllauer L, Gruber P, Sebinger D et al. (2001). Mutations in apoptosis genes: a pathogenetic factor for human disease. *Mutation Research* 488: 211–31.
21. Gross A, McDonnell JM, Korsmeyer SJ. (1992). *BCL-2* family members and the mitochondria in apoptosis. *Genes Dev* (13): 1899 – 1911.
22. Nagata S. (2004). Early work on the function of CD95, an interview with Shige Nagata. *Cell Death and Differentiation* 11: S23 – 27. doi: 10.1038/sj.cdd.4401453
23. Pawlowski, Kraft AS. (2000). BAX-induced apoptosis cell death. *PNAS* 97 (2): 529 – 531.
24. Oda E, Ohki R, Murasawa H et al. (2000). Noxa, a BH3-only family member of the Bcl-2 family and candidate mediator of p53-induced apoptosis. *Science* 288: 1053 – 1058.
25. Miller LK. (1999). An exegesis of IAPs: Salvation and surprises from BIR motifs. *Trends Cell Biol* 9: 323 – 328.
26. Biliran H Jr, Wang Y, Banerjee S et al. (2005). Overexpression of cyclin D1 promotes tumour cell growth and confers resistance to cisplatin-mediated apoptosis in an elastase-myc transgene-expressing pancreatic tumour cell line. *Clin Cancer Res* 1511 (16): 6075 – 6086.
27. Ahn BK, Kim SH, Paik SS, Lee KH. (2016). Loss of APAF-1 expression is associated with early recurrences in stage I, II and III colorectal cancer. *Langenbeck's Archives of Surgery* 401 (8): 1203 – 1210.
28. Bagby G. (2018) Recent advances in understanding haematopoiesis in Fanconi anaemia. *F1000Research* 7: 105. doi: 10.12688/f1000research.13213.1.
29. Rieux-Laucat FR, Magerus-Chatinet A. (2010). Autoimmune lymphoproliferative syndrome: A multifactorial disorder. *Haematologica* 95 (11): 1805 – 1807.
30. Mitroulis I, Kambas K, Ritis K. (2013). Neutrophils, IL1- β and gout: Is there a link? *Semin Immunopathol*. doi 10.1007/s00281-013-0361-0.
31. Verzola D, Ratto E, Villaggio B et al. (2014). Uric acid promotes apoptosis in human proximal tubule cells by oxidative stress and the activation of NADPH oxidase NOX4. *PLoS One* 9 (12): e115210. doi: 10.1371/journal.pone.0115210.
32. Soung YH, Lee JW, Kim SY et al. (2005). CASPASE -8 gene inactivated by somatic mutations in gastric carcinomas. *Cancer Res* 65 (3): 815 – 821.
33. Kurokawa M, Ito T, Yang CH, Zhao C et al. (2013). Engineering a BCR-ABL-activated caspase for the selective elimination of leukemic cells. *PNAS* 110 (6): 2300 – 2305.
34. Stone S, Ho Y, Li X et al. (2016). Dual role of integrated stress response in medulloblastoma tumorigenesis. *Oncotarget* 7 (39): 64124 – 64135. doi: 10.18632/oncotarget.11873.
35. Baens, B, Maes A, Steyls K et al. (2000). The product of the t(11;18), an API2-MLT fusion, marks nearly half of gastric MALT type lymphomas without large cell proliferation. *Am J Pathol* 156: 1433 – 1439.
36. Wang L, Lin S, Yi D et al. (2017). Apoptosis, expression of PAX3 and p53 and caspase signal in fetuses with neural tube defects. *Birth Defects Res* 109 (19): 1596 – 1604. doi: 10.1002/bdr2.1094.
37. Ahn EJ, Yum MS, Kim EH et al. (2017). Genotype- phenotype correlation of SMN1 and NAIP deletions in Korean patients with spinal muscular atrophy. *J Clin Neurol* 13-1: 27 – 31. doi: 10.3988/jcn.2017.13.1.27
38. Wu H, Zhao Z, Liu J et al. (2018). Long noncoding RNAMeg3 regulates cardiomyocyte apoptosis in myocardial infarction. *Gene Ther*. doi: 10.1038/s41434-018-0045-4.

39. Aljakna A, Fracasso T, Sabatasso S. (2018). Molecular tissue changes in early myocardial ischemia: from pathophysiology to the identification of new diagnostic markers. *Int J Legal Med* 132: 425 – 438. Retrieved from <https://doi.org/10.1007/s00414-017-1750-z>.
40. Laguna RK, Creasey EA, Li Z et al. (2006). *Legionella pneumophila*-translocated substrate that is required for growth within macrophages and protection from host cell death. *Proc Natl Acad Sci USA* 103: 18745 – 18750. doi: 10.1073/pnas.0609012103.
41. Ying S, Seiffert BM, Häcker G, Fischer SF. (2005). Broad degradation of proapoptotic proteins with the conserved Bcl-2 homology domain 3 during infection with *Chlamydia trachomatis*. *Infect Immun* 73: 1399 – 1403. doi: 10.1128/IAI.73.3.1399-1403.2005.
42. Velmurugan KB, Chen JL, Miller S et al. (2007). *Mycobacterium tuberculosis* *nuoG* is a virulence gene that inhibits apoptosis of infected host cells. *PLoS Pathog* 3: e110. doi: 10.1371/journal.ppat.0030110.
43. Parthasarathy G, Philipp M. (2012). Review: Apoptotic mechanisms in bacterial infections of the central nervous system. *Front Immunol* 3 (306): 1 – 13. doi: 10.3389/fimmu.2012.00306.
44. Barichievy S, Naidoo J, Boullé M et al. (2018). MAP2K1/ERK2 inhibitors as a novel therapeutic intervention strategy for HIV-1 infection in macrophages. *Front Cell Infect Microbiol* 8: 263. doi: 10.3389/fcimb.2018.00263
45. Kawahara R, Rosa-Fernandes L, Santos AF et al. (2018). Integrated proteomics reveals apoptosis-related mechanisms associated with placental malaria. *Mol Cell Proteomics* in Press Manuscript RA118.000907.
46. Mankouri J, Dallas ML, Hughes ME et al. (2009). Suppression of a proapoptotic K⁺ channel as a mechanism for hepatitis C virus persistence. *Proc Natl Acad Sci USA* 106 (37): 15903 – 15908. doi: 10.1073/pnas.0906798106.
47. Schaaf K, Smith SR, Duverger A et al. (2017). *Mycobacterium tuberculosis* exploits the PPM1A signalling pathway to block host macrophage apoptosis. *Scientific Reports* 7 (42101): 1 – 16. doi: 10.1038/srep42101.
48. de Souza Aarao TL, de Sousa JR, Falcao ASC et al. (2018). Nerve growth factors and pathogenesis of leprosy: Review and Update. *Front Immunol*. Retrieved from <https://doi.org/10.3389/fimmu.2018.00939>.
49. Du P, Li S, Ojcius DM et al. (2018). A novel Fas-binding outer membrane protein and lipopolysaccharide of *Leptospira interrogans* induce macrophage apoptosis through the Fas/FasL-Caspase-8/-3 pathway. *Emerging Microbes & Infections* 7: 1 – 17.
50. Chen X, Wang T, Yang D et al. (2013). Expression of the IAP protein family acts cooperatively to predict prognosis in human bladder cancer patients. *Oncology letters* 5: 1278 – 1284. doi: 10.3892/ol.2013.
51. Hennessy EJ, Adam A, Aquila BM, Castriotta LM et al. (2013). Discovery of a novel class of dimeric Smac mimetics as IAP antagonists resulting in a clinical candidate for the treatment of cancer. *Journal of Medicinal Chemistry* 56 (24): 9897 – 9919.
52. Yang L, Cao Z, Yan H, Wood WC. (2003). Coexistence of high levels of apoptotic signalling and inhibitor of apoptosis proteins in human tumour cells: implications for cancer specific therapy. *Cancer Res* 63(20): 6815 – 6824.
53. Wang Y, Zhang H, Liu C et al. (2013). Enhancement of survivin-specific-antitumor immunity by adenovirus prime protein-boost immunity strategy with DDA/MPL adjuvant in a murine melanoma model. *International Immunopharmacology* 17: 9 – 17. <https://doi.org/10.1016/j.intimp.2013.04.015>.
54. Baig S, Seevasant I, Mohamad J et al. (2016). Potential of apoptotic pathway-targeted cancer therapeutic research: Where do we stand? *Cell Death & Disease* 7: e2058.
55. Palacios C, Yerbes R, Sanchez-Perez T et al. (2014). The long and winding road to cancer treatment: the TRAIL system. *Current Pharm Des* 20 (17): 2819 – 2833.
56. Ebrahim AS, Sabbagh H, Liddane A et al. (2016). Haematologic malignancies: Newer strategies to counter the BCL-2 protein. *J Cancer Res Clin Oncol* 142: 2013 – 2022. doi: 10.1007/s00432-016-2.
57. Benjamin LW, Wen XD, Hartmut J. (2017). Caspase inhibitors for the treatment of liver disease: friend or foe? *Expert Rev Gastroenterol Hepatol* 11 (5): 397–399.
58. Chanel Li Keoni, Brown TL. (2015). Inhibition of apoptosis and efficacy of pancaspase inhibitor QVD-OPh in models of human diseases. *J Cell Death* (8): 1 – 7.
59. Maj E, Papiernik D, Wietrzyk J. (2016). Angiogenic cancer treatment: the great discovery and greater complexity. (Review) *Int J Oncol* 49 (5): 1773 – 1784.